

**REVIEW ARTICLE****Mucoadhesive Nasal Drug Delivery System**

**Mhetre R.M.\*, Gujrathi D.S. , Shrisundar N.S., Holikatti S.S., Lad G.C.,  
Deshmane P.S., Swami M.M.**

**Gandhi Natha Rangji College of D.Pharmacy, Solapur**

**Corresponding author: Rani Mallinath Mhetre, At Post Doddi, Tq. South Solapur, Dist. Solapur,  
Maharashtra- 413006, India**

**ABSTRACT**

Nasal drug delivery system has shown great attraction in the past years to optimized therapeutic effect of drug, due to high permeability of nasal epithelial membrane so that rapid absorption of drug is possible, as compared to other non-invasive routes Nasal drug delivery system provides easy application of drug, with the possibility of self administration by removing the chance of unwanted painful condition associated with injection form of drug delivery .in this review ,the importance of nasal drug delivery system along with its advantages over other routes of administration is enlisted.

Keywords: microspheres, mucociliary clearance ,mucoadhesion.

**INTRODUCTION****INTRODUCTION:**

Oral drug delivery is the most desirable route for drug administration whenever systemic effects are intended. Therefore, it is not surprising that the prediction of human oral bioavailability of new drug candidates is currently targeted from the earliest stages of drug discovery and development programmes <sup>1, 2</sup>. However, although the oral route remains the most popular for systemic drug administration but low oral bioavailability of some compounds has prompted the search of more effective routes for their systemic delivery <sup>3</sup>.

Nasal drug delivery system has shown great attraction in the past years to optimized therapeutic effect of drug, due to high permeability of nasal epithelial membrane so that rapid absorption of drug is possible, as compared to other non-invasive routes<sup>1,2</sup> Nasal drug delivery system provides easy application of drug, with the possibility of self administration by removing the chance of unwanted painful condition associated with injection form of drug delivery. Furthermore, lipophilic and low molecular weight drugs can easily penetrate through nasal mucosa with less degradation. Fast absorption can be achieved due to large absorption surface area and high vascularisation. Nasal route can be used as an alternative to parenteral in case of emergency therapy.<sup>3,4</sup> Nasal drug delivery system is a potential route for direct delivery of drug to the central nervous system through olfactory region bypassing hepatic first pass metabolism.<sup>5,6</sup> Side by side nasal drug delivery system has some limitations like large dose cannot be administered by this route conveniently due to administrative problems. Administration of solid formulation is quite difficult by nasal route.<sup>5</sup> Fast clearance of the administered formulation

occurs from the nasal cavity as the result of mucociliary clearance causes poor absorption of drug.<sup>7</sup> These difficulties of nasal route can be minimized by utilization of various kinds of mucoadhesive polymers in the formulation. These polymers can effectively increase the retention time with improved permeation enhancing effect. In some research these polymers also possess the controlled release of drug. A variety of polymers have been discovered which includes, synthetic as HPMC, HEC, Chitosan, Carbopol and natural as gelatin, albumin, starch. Utility of synthetic polymers are associated with large numbers of risk such as high cost, toxicity, environmental pollution during synthesis, non renewable sources, side effects and poor patient compliance.<sup>8</sup> These limitations of synthetic polymers may be avoided by utilization of natural polymers as they are biodegradable, chemically inert, less expensive, nontoxic, and widely available.<sup>1,8</sup> Natural products are now accepted worldwide due to their biodegradability, which leads low chance of risk during uses.

Nasal administration can therefore be used as an alternative to oral administration of for example tablets and capsules if a fast effect is desired or if the drug is extensively degraded in the gut or liver. Therapy through intranasal administration has been an accepted form of treatment in the Ayurvedic system of Indian medicine. Historically, nasal drug delivery system has received interest since ancient times. Nasal administration can be used to deliver drugs for either local or systemic effect. Locally acting drugs are for example decongestants and allergy treatments. Examples of systemically active drugs available as nasal sprays are migraine drugs, nicotine replacement and hormone treatments. In order to formulate a nasal formulation with desirable performance and commercial attributes, the drug properties, delivery system and nasal physiology should all be considered and understood from the early stages of a product development. It is advisable to focus on maximizing the residence time and ensuring an efficient absorption of drug. A successful nasal formulation program involves detailed consideration of the interactions between formulation composition, device design, delivery system and the patient's pathological condition. If a nasal formulation is delivered to the target site of absorption (turbinates), benefits can be gained from increased absorption and/or decreased dosage requirements. There may also be a reduction of taste of the drug because of minimum or reduced swallowing of the administered drug. Currently, tip aperture design pumps are available to administer formulations in an upward direction. Because the turbinates are located at the sides of the nostrils, the entire dose volume cannot be administered to the target site of absorption. This also leads to swallowing of part of the dose. It may be possible to design a side aperture pump to direct the entire dose volume directly to the absorption site, the turbinate's, for more efficient (target) nasal delivery. Nasal sprays for local effect are quite common. Several anti-migraine drugs are also currently administered by nasal administration because a fast effect is desired and oral administration can be prohibited by nausea. Peptide drugs (hormone treatments) are also available as nasal sprays, in this case to avoid drug degradation after oral administration. The peptide analogue desmopressin is, for example, available for both nasal and oral administration.

The bioavailability of the commercial tablet is 0.1% while that of the nasal spray is 3-5% according to the SPC (summary of product characteristics). Other potential drug candidates for nasal administration include anaesthetics, antiemetics and sedatives that all benefit from a fast onset of effect.<sup>5</sup>

Nasal drug delivery is a useful delivery method for drugs that are active in low doses and show no minimal oral bioavailability. The nasal route circumvents hepatic first pass elimination associated with the oral delivery: it is easily accessible and suitable for self-medication

Currently, two classes of nasally delivered therapeutics are on the market. The first one comprises low molecular weight and hydrophobic drugs for the treatment of the nasal mucosa and sinus, including decongestants, topical steroids, antibiotics and other (OTC) products. The second class encompasses a few drugs, which have sufficient nasal absorption for displaying systemic effects. Important candidates are the compounds, generally administered by injection and hardly absorbed after oral administration, due to their instability in gastrointestinal tract, poor absorption properties, and their rapid and extensive biotransformation. Therefore, nasal delivery is promising alternative route for the administration of peptides and protein drugs in particular.

Nasal mucosa has been considered as a potential administration route to achieve fast and higher level of drug absorption because it is permeable to more compounds than the gastrointestinal tract due to lack of pancreatic and gastric enzymatic activity, neutral pH of the nasal mucus and less dilution by gastrointestinal contents<sup>8</sup>. In recent years many drugs have been shown to achieve better systemic bioavailability through nasal route than by oral administration. Nasal therapy, has been recognized form of treatment in the Ayurvedic systems of Indian medicine, it is also called “NASAYA KARMA”<sup>9</sup>. Intranasal drug delivery – which has been practiced for thousands of years, has been given a new lease of life. It is a useful delivery method for drugs that are active in low doses and show no minimal oral bioavailability such as proteins and peptides<sup>10</sup>. One of the reasons for the low degree of absorption of peptides and proteins via the nasal route is rapid movement away from the absorption site in the nasal cavity due to the mucociliary clearance mechanism<sup>11</sup>. The nasal route circumvents hepatic first pass elimination associated with the oral delivery. IN non-invasive, essentially painless, does not require sterile preparation, and is easily and readily administered by the patient or a physician, e.g., in an emergency setting. Furthermore, the nasal route may offer improved delivery for “non-Lipinski” drugs<sup>12</sup>. Drug candidates ranging from small metal ions to large macromolecular proteins have been tested in various animal models<sup>9</sup>.

### **ADVANTAGES<sup>13</sup>**

Drug degradation that is observed in the gastrointestinal tract is absent.

- Hepatic first pass metabolism is avoided.
- Rapid drug absorption and quick onset of action can be achieved.
- The bioavailability of larger drug molecules can be improved by means of absorption enhancer or other approach.
- The nasal bioavailability for smaller drug molecules is good.
- Drugs that are orally not absorbed can be delivered to the systemic circulation by nasal drug delivery.
- Studies so far carried out indicate that the nasal route is an alternate to parenteral route, especially, for protein and peptide drugs.
- Convenient for the patients, especially for those on long term therapy, when compared with parenteral medication.
- Drugs possessing poor stability in g.i.t. fluids are given by nasal route.
- Polar compounds exhibiting poor oral absorption may be particularly suited for this route of delivery

### **LIMITATIONS<sup>14,15</sup>**

- The histological toxicity of absorption enhancers used in nasal drug delivery system is not yet clearly established.

- Relatively inconvenient to patients when compared to oral delivery systems since there is a possibility of nasal irritation.
- Nasal cavity provides smaller absorption surface area when compared to GIT.
- There is a risk of local side effects and irreversible damage of the cilia on the nasal mucosa, both from the substance and from constituents added to the dosage form.
- Certain surfactants used as chemical enhancers may disrupt and even dissolve membrane in high concentration.
- There could be a mechanical loss of the dosage form into the other parts of the respiratory tract like lungs because of the improper technique of administration.

### **MUCOADHESIVE DRUG DELIVERY SYSTEM**

Mucoadhesive drug delivery system are delivery system which utilizes the property of bioadhesion of certain polymers which become adhesive on hydration and can be used for targeting a drug to a particular region of the body for extended periods of time. The term “mucoadhesion” was coined for the adhesion of the polymers with the surface of the mucosal layer<sup>83</sup>. Bioadhesion is a phenomenon in which two materials at least one of which is biological and are held together by means of interfacial forces. The attachment could be between an artificial material and biological substrate such as adhesion between polymer and a biological membrane in case of polymer attached to the mucin layer of mucosal tissue. The term mucoadhesion is used when the mucosal layer lines a number of regions of body including a gastrointestinal tract, urogenital tract, the airways, the ears, nose and eye. These represent potential sites for attachment of bioadhesive system and hence the mucoadhesive drug delivery system could be designed for buccal, oral, vaginal, rectal, nasal and ocular route of administration.

The nasal route of drug administration constitutes the one of the rare and recent and preferred means of drug delivery to systemic circulation of body. However nasal administration of most of the drugs in liquid dosage forms has short-term limitations due to their inability to restrain and localize at site of administration. Microspheres constitute an important part of these particulate drug delivery systems by virtue of their small size and efficient carrier capacity. Microspheres are the carrier linked drug delivery system in which particle size is ranges from (1-1000  $\mu\text{m}$ ) range in diameter having a core of drug and entirely outer layers of polymers as coating material. However, the success of these microspheres is limited due to their short residence time at site of absorption. It would, therefore be advantageous to have means for providing an intimate contact of the drug delivery system with the absorbing membrane<sup>82</sup>. This can be achieved by coupling bioadhesion characteristics to microspheres and developing bioadhesive microspheres. Bioadhesive microspheres have advantages like efficient absorption and enhanced bioavailability of the drugs due to a high surface to volume ratio, a much more intimate contact with the mucus layer and specific targeting of drugs to the absorption site.

### **MECHANISM OF MUCOADHESION**

A complete understanding of how and why certain macromolecules attach to a mucus surface is not yet available, but a few steps involved in the process are generally accepted, at least for solid systems. Several theories have been proposed to explain the fundamental mechanism of adhesion such as<sup>16</sup>

- 1) Electronic theory
- 2) Absorption theory
- 3) Diffusion theory

- 4) Wetting theory
- 5) Cohesive theory

A General Mechanism of Mucoadhesion Drug Delivery system is shown in Figure 1.

**Electronic theory**

According to this theory, electron transfers occur upon contact of adhesive polymer with a mucus glycoprotein network because of difference in their electronic structures. This results in the formation of electrical double layer at the interface e.g. Interaction between positively charged polymers chitosan and negatively charged mucosal surface which becomes adhesive on hydration and provides an intimate contact between a dosage form and absorbing tissue.

**Absorption theory**

According to this theory, after an initial contact between two surfaces, the material adheres because of surface force acting between the atoms in two surfaces. Two types of chemical bonds resulting from these forces can be distinguished as primary chemical bonds of covalent nature and Secondary chemical bonds having many different forces of attraction, including electrostatic forces, Vander Walls forces, hydrogen and hydrophobic bonds.

**Diffusion theory**

According to this theory, the polymer chains and the mucus mix to a sufficient depth to create a semi permanent adhesive bond. The exact depth to which the polymer chain penetrates the mucus depends on the diffusion coefficient and the time of contact. The diffusion coefficient in terms depends on the value of molecular weight between crosslinking and decreases significantly as the cross linking density increases.

**Wetting theory**

The wetting theory postulates that if the contact angle of liquids on the substrate surface is lower, then there is a greater affinity for the liquid to the substrate surface. If two substrate surfaces are brought in contact with each other in the presence of the liquid, the liquid may act as an adhesive among the substrate surface.

**Cohesive theory**

The cohesive theory proposes that the phenomena of bioadhesion are mainly due to intermolecular interaction amongst like molecule. Based upon the above theories, the process of bioadhesion can broadly be classified into two categories namely chemical (electron and absorption theory) and physical (wetting, diffusion and cohesive theory).

**POLYMERS USED IN MUCOADHESIVE DRUG DELIVERY SYSTEM**

Mucoadhesive polymers are water-soluble and water insoluble polymers, which are swellable networks, jointed by cross-linking agents. These polymers possess optimal polarity to make sure that they permit sufficient wetting by the mucus and optimal fluidity that permits the mutual adsorption and interpenetration of polymer and mucus to take place. Following types of polymers are used in mucoadhesive drug delivery system:

- 1) Hydrophilic polymers  
E.g. Anionic polyelectrolytes - poly (acrylic acid) and carboxymethyl cellulose  
Cationic polyelectrolyte – chitosan  
Non-ionic polymers - poloxamer, hydroxypropyl methyl cellulose, methyl cellulose, Poly (vinyl alcohol) and poly (vinyl pyrrolidone)
- 2) Hydrogels  
E.g. poly acrylic acid
- 3) Thiolated polymers  
E.g. Chitosan- iminothiolane, poly (acrylic acid)–cysteine, poly (acrylic acid)–homocysteine, chitosan–thioglycolic acid, chitosan–thioethylamidine, alginate–cysteine, poly (methacrylic acid)–cysteine and sodium carboxymethylcellulose–cysteine,<sup>80</sup>
- 4) Lectin based polymers

### **IDEAL CHARACTERISTICS OF AN MUCOADHESIVE POLYMER**

1. The polymer and its degradation products should be nontoxic and nonabsorbable from the GIT.
2. It should be nonirritant to the mucous membrane.
3. It should preferably form a strong noncovalent bond with the mucin-epithelial cell surfaces.
4. It should adhere quickly to most tissue and should possess some site-specificity.
5. It should allow daily incorporation to the drug and offer no hindrance to its release.
6. The polymer must not decompose on storage or during the shelf life of the dosage form.
7. The cost of polymer should not be high so that the prepared dosage form remains competitive

### **METHOD OF PREPARATION**

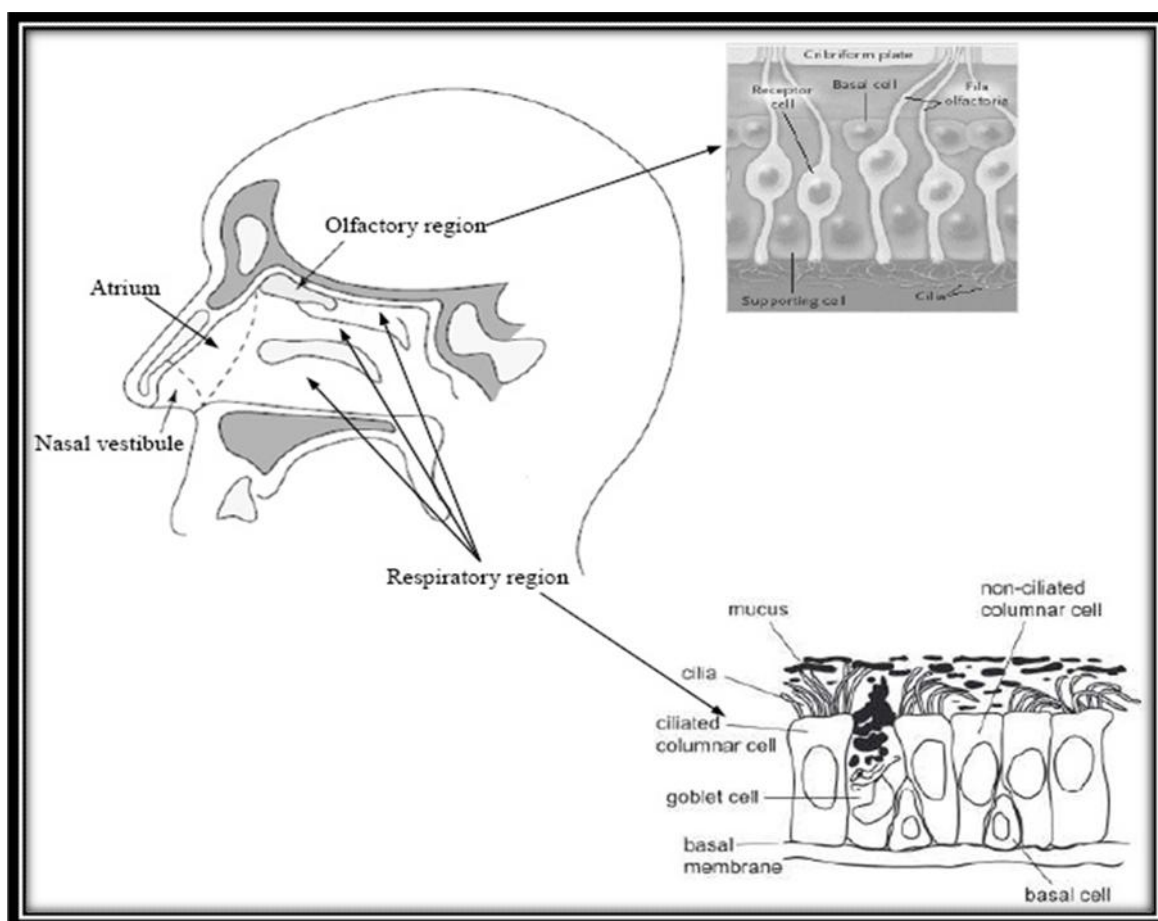
- Preparation of Microspheres by Thermal cross-linking
- Preparation of Microspheres by Glutaraldehyde crosslinking
- Preparation of microspheres by Tripolyphosphate
- Preparation of Microspheres by Emulsification and Ionotropic gelation by NaOH
- Preparation of Ethyl cellulose Microspheres
- Spray Drying
- Solvent Evaporation
- Wet Inversion Technique
- Complex Coacervation
- Hot Melt Microencapsulation

### **ANATOMY & PHYSIOLOGY OF NASAL CAVITY**

The nasal cavity is divided into two halves by the nasal septum and extends posterior to the nasopharynx, while the most anterior part of the nasal cavity, the nasal vestibule, opens to the face through the nostril. The nasal cavity consists three main regions are nasal vestibule, olfactory region and respiratory region. The surface area in the nose can be enlarged about 150cm by the lateral walls of the nasal cavity includes a folded structure, it is a very high surface area compared to its small volume. This folded structure consists of three turbinates: the superior, the median and the inferior<sup>17</sup>. The main nasal airway having the narrow passages,



usually it has 1-3mm wide and these narrow structures are useful to nose to carry out its main functions. The nasal cavity is covered with a mucous membrane which can be divided into two areas; non olfactory and olfactory epithelium, in this non-olfactory area includes the nasal vestibule which is covered with skin-like stratified squamous epithelium cells, where as respiratory region, which has a typical airways epithelium covered with numerous microvilli, resulting in a large surface area available for drug absorption and transport (Sarkar 1992). In this way the mucus layer is propelled in a direction from the anterior to-towards the posterior part of the nasal cavity. The goblet cells are present in the mucus membrane which covers the nasal turbinate and the atrium; it secretes the mucus as mucus granules which are swelling in the nasal fluid to contribute to the mucus layer. The mucus secretion is composed of about 95% water, 2 % mucin, 1% salts, 1% of other proteins such as albumin, immunoglobulins, lysozyme and lactoferrin, and about 1% lipids (Kaliner et al., 1984). The mucus secretion gives immune protection against inhaled bacteria and viruses. It also performs a number of physiological functions. <sup>1</sup> It covers the mucosa, and physically and enzymatically protects it. <sup>2</sup> The mucus has waterholding capacity. <sup>3</sup> It exhibits surface electrical activity. <sup>4</sup> It permits efficient heat transfer. <sup>5</sup> It acts as adhesive and transports particulate matter towards the nasopharynx.



**Fig.2. Anatomy and Physiology of nasal cavity**

There is mainly four parts in the nasal cavity called vestibule, atrium, respiratory region and olfactory region. Each part distinguishes from one other due to their specific characteristic, function and permeability.

#### **Nasal vestibule**

Nasal vestibule is the most anterior part of the nasal cavity, just inside the nostrils, and presents an area about 0.6 cm<sup>2</sup><sup>18</sup>. Here, there are nasal hairs, also called vibrissae, which filter the inhaled particles. Histologically, this nasal portion is covered by a stratified squamous and keratinized epithelium with sebaceous glands<sup>18,19,20</sup>. These nasal vestibular characteristics are desirable to afford high resistance against toxic environmental substances but, at the same time, the absorption of substances including drugs becomes very difficult in this region<sup>21</sup>.

#### **Atrium**

Atrium is the intermediate area between nasal vestibule and respiratory region. Its anterior section is constituted by a stratified squamous epithelium and the posterior area by pseudostratified columnar cells presenting microvilli<sup>19,20</sup>.

#### **Respiratory region**

The nasal respiratory region, also called conchae, is the largest part of the nasal cavity and it is divided in superior, middle and inferior turbinates which are projected from the lateral wall. These specialized structures are responsible for humidification and temperature regulation of inhaled air. Between them there are spaces, called meatus, which are passageways where airflow is created to assure a close contact of the inhaled air with the respiratory mucosal surface. The inferior and middle meatus receive nasolacrimal ducts and paranasal sinuses which are air-filled pockets located inside the bones of the face and around the nasal cavity<sup>22</sup>. The nasal respiratory mucosa, considered the most important section for delivering drugs systemically, is constituted by the epithelium, basement membrane and lamina propria. The nasal respiratory epithelium consists of pseudostratified columnar epithelial cells, goblet cells, basal cells and mucous and serous glands<sup>19,20,23</sup>. Many of the epithelial cells are covered on their apical surface with microvilli and the major part of them also has fine projections, called cilia<sup>42</sup>. Actually, microvilli are important to enhance the respiratory surface area, while cilia are essential to transport the mucus toward the nasopharynx. Under physiological conditions, nasal epithelium is covered with a thin mucus layer produced by secretory glands and goblet cells. These ones secrete granules filled with mucin, a glycoprotein that determines the viscosity of the mucus. The nasal mucus layer is only 5 µm thick and it is organized in two distinct layers: an external, viscous and dense, and an internal, fluid and serous. Overall, nasal mucus layer consists of 95% of water, 2.5-3% of mucin, and 2% of electrolytes, proteins, lipids, enzymes, antibodies, sloughed epithelial cells and bacterial products<sup>24,25</sup>. Nasal mucus is indispensable for several physiological functions, such as humidification and warming of the inhaled air, and also offers physical and enzymatic protection of the nasal epithelium against several foreign compounds, including drugs. The protective action results of the adhesive characteristics of mucus to attract inhaled particles or pathogens, which are removed towards the nasopharynx by nasal MCC<sup>26</sup>. The presence of mucin in the nasal mucus layer is crucial because it may trap large molecular weight drugs, such as peptides and proteins<sup>27</sup>. The basal cells that exist in the epithelium are progenitors of other cell-types and lie on a thickened layer of collagen called basement membrane. Beneath of it, there is the lamina propria which is richly supplied with blood vessels, including many very permeable fenestrated capillaries, nerves, glands and immune cells. The last



ones produce immunoglobulin A antibodies that confer immunological protection against bacteria and virus <sup>28</sup>.

### Olfactory region

The olfactory region is located in the roof of the nasal cavity and extends a short way down the septum and lateral wall <sup>41</sup>. Its neuroepithelium is the only part of the CNS that is directly exposed to the external environment <sup>29</sup>. Similarly to the respiratory epithelium, the olfactory one is also pseudostratified but contains specialized olfactory receptor cells important for smell perception <sup>29,30</sup>. In this area there are also small serous glands (glands of Bowman) producers of secretions acting as a solvent for odorous substances <sup>30</sup>.

**Table: 1- A feature of specific parts of nasal cavity:**

Nasal parts	Characteristics	Function	Permeability	Surface area	vascularization
Vestibule	Keratinized and stratified squamous epithelial cells with nasal hairs	Support and protection	Poor	~0.6 cm <sup>2</sup>	low
Atrium	Stratified squamous cells and pseudostratified cells	Support	Reduced	NF	low
Respiratory region	Columnar ciliated cells, columnar non ciliated cells, goblet	Support, muciliary clearance and Mucus secretion	Good	~130 cm <sup>2</sup>	Very high
Olfactory region	Sustentacular cells, olfactory receptor cells, and basal cells	Support and olfaction Perception	Direct access to CNS	~15 cm <sup>2</sup>	High

### MECHANISM OF NASAL ABSORPTION

First mechanism involves paracellular route of transport, which is a passive process of absorption through nasal route. Hydrophilic drugs transport through this route. The drugs with molecular

weight greater than 1000 daltons show poor bioavailability<sup>31</sup>. Second mechanism involves transcellular process. Lipophilic drugs transport through this route. It is an active route of transport.

### **1. First mechanism**

It involves an aqueous route of transport, which is also known as the paracellular route but slow and passive. There is an inverse log-log correlation between intranasal absorption and molecular weight of water-soluble compounds. The molecular weight greater than 1000 Daltons having drugs shows poor bioavailability.

### **2. Second mechanism**

It involves transport through a lipoidal route and it is also known as the transcellular process. It is responsible for the transport of lipophilic drugs that show a rate dependency on their lipophilicity. Drug also cross cell membranes by an active transport route via carrier-mediated means or transport through the opening of tight junctions.

Intranasal Drug Delivery is mainly used for two purposes i.e. for mainly systemic delivery and for local delivery. To assess the therapeutic viability of intranasal drug delivery several approaches should be considered, attending, specifically, to the nature of pathologic condition (acute or chronic) and intended effects of drug treatment (local, systemic or at CNS). Indeed, for acute disease conditions, the advantages afforded by intranasal drug delivery in terms of patient comfort and compliance may not be much relevant when compared with drug delivery by parenteral route. In contrast, this is particularly important to treat clinical or chronic condition.<sup>3</sup>

### **Local delivery**

Intranasal administration of medicines is the natural choice for the treatment of topical nasal disorders. Among the most common examples are antihistamines and corticosteroids for rhinosinusitis, and nasal decongestants for cold symptoms (Table 2). In these cases, intranasal route is the primary option for drug delivery because it allows a rapid symptom relief with a more favourable adverse-event profile than oral or parenteral routes. In fact, relatively low doses are effective when administered topically<sup>18</sup>, minimizing simultaneously the potential of systemic toxic effects. Recently, for instance, topical antibiotherapy has been considered in chronic rhinosinusitis in an attempt to eradicate biofilm bacteria, often resistant to systemic treatment, and still avoiding systemic toxicity.

### **Systemic delivery**

The intranasal administration is an effective way to systemically deliver drugs as an alternative to oral and intravascular routes. Actually, it seems to present fast and extended drug absorption<sup>32</sup>, and it has been supported by many studies planned to compare intranasal drug delivery against oral and parenteral administration (Figure 2)<sup>33,34,35</sup>. Consequently, the number of drugs administered as nasal formulations intended to achieve systemic effects has widely increased. Some prominent examples include analgesics (morphine)<sup>18,36,37</sup>, cardiovascular drugs as propranolol<sup>37</sup> and carvedilol<sup>38</sup> hormones such as levonorgestrel<sup>39</sup>, progesterone<sup>38</sup> and, hormones insulin<sup>43,44,45,46</sup> anti-inflammatory agents as indomethacin<sup>46,47</sup> and ketorolac<sup>48,49</sup>, and antiviral drugs (acyclovir)<sup>40,41,42</sup>. Actually, there are some examples already available in the market (Table 2). These include, for instance, zolmitriptan and sumatriptan for the treatment of migraine and cluster headaches.

**Table 2: Examples of nasal formulations commercially available after prescription** <sup>18,50,51,52</sup>  
**Local Delivery**

Drug	Brand	Main Expeints	Supplier	Main Indications
Azelastine	Astelin	Benzalkonium chloride, edetate disodium, hypromellose	Meda Pharmaceuticals	
Beclometasone	Beconase	Microcrystalline cellulose, carboxymethyl cellulose sodium, benzalkonium chloride	GlaxoSmithKline	
Budesonide	Rhinocort	Microcrystalline cellulose, carboxymethyl cellulose sodium, dextrose anhydrous	AstraZeneca	
Levocabastine	Livostin	Benzalkonium chloride, edetate disodium, disodium phosphate	Jansen-Cilag	Management/treatment of symptoms of seasonal and perennial rhinosinusitis
Mometasone	Nasonex	Microcrystalline cellulose, carboxymethylcellulose sodium, benzalkonium chloride	Schering-Plough	
Olapatadine	Patanase	Benzalkonium chloride, dibasic sodium phosphate, edetate disodium	Alcon Laboratories	
Sodium cromoglicate	Nasalcrom	Benzalkonium chloride, edetate disodium	Sanofi-Aventis	
Triamcinolone acetonide	Nasacort	Microcrystalline cellulose, carboxymethylcellulose sodium, polysorbate 80	Sanofi-Aventis	
Mupirocin	Bactroban	Paraffin and a mixture of glycerine esters (Softisan 649)	GlaxoSmithKline	Eradication of nasal staphylococci

## Systemic Delivery

**Table 3 nasal; formulations used for the purpose of systemic delivery**

Estradiol	Aerodiol	Methylbetadex, sodium chloride	Servier laboratories	Hormone replacement therapy
Nicotine	Nicotrol NS	Disodium phosphate, sodium dihydrogen phosphate, citric acid	Pfizer	Smoking cessation
Cyanocobalamin	Nascobal	Sodium citrate, citric acid, benzalkonium chloride	Strativa pharmaceuticals	Vitamin B12 deficiency
Desmopressin	Desmospray	Sodium chloride, citric acid, benzalkonium chloride	Ferring Pharmaceuticals	Control of dehydration in diabetes insipidus
Oxytocin	Syntocinon	Citric acid, chlorobutanol, sodium chloride	Novartis	Labour induction; lactation stimulation
Salmon calcitonin	Miacalcin	Sodium chloride, benzalkonium chloride, hydrochloric acid	Novartis	Treatment of postmenopausal osteoporosis
Buserelin	Suprefact	Sodium hydroxide, sodium chloride, sodium dihydrogen phosphate	Sanofi-Aventis	Treatment of prostate cancer
Nafarelin	Synarel	Benzalkonium chloride, glacial acetic acid	Roche Laboratories	Management of endometriosis
Sumatriptan	Imigran	Potassium dihydrogen cluster headaches phosphate, dibasic sodium phosphate anhydrous	GlaxoSmithKline	Treatment of migraine and Sumatriptan Imigran Potassium dihydrogen cluster headaches
Fentanyl	Instany	Sodium dihydrogen phosphate dehydrate, disodium phosphate dehydrate	Nycomed Pharma	
Butorphanol	Stadol NS	Sodium chloride, citric acid, benzethonium chloride	Bristol-Myers Squibb	Pain management
Live attenuated influenza vaccine	FluMist	Monosodium glutamate, hydrolyzed porcine gelatin, arginine, dibasic potassium phosphate, monosodium phosphate, gentamicin sulfate	MedImmune, Inc	Flu prevention

**BARRIERS TO NASAL ABSORPTION**

Nasal drug delivery system is considered has a profitable route for the formulation scientist because it has easy and simple formulation strategies. Intra-nasally administered drug products therapeutic efficacy and toxicities are influenced by number of factors. Following factors are the barriers to the absorption of drugs through nasal cavity.

**Table 4 :Barriers to nasal absorption :**

Nasal Barriers	Factors to be considered
1) <b>Physiological barriers</b>	
a) Nasal mucus	a)Viscosity ph of mucous drug and dosage form interaction
b) Nasal epithelial barrier	b)molecular weight, ionization constant and mode of transport
c) Mucociliary clearance	c) nasal residential time and nature of dosage form
d) Pathophysiology	d)volume of nasal secretion and permeability of nasal epithelium
e) Nasal metabolism	e) nature of the molecules (e.g. proteins and peptides)
f) Efflux transport system	f) nature of drug molecules and duration of therapy
2) <b>Physicochemical barriers</b>	
a) Drug solubility and dissolution	a) Nature of the dosage form , dose , pka, and polymorphism
b) Molecular weight and size	b) Less bioavailabitliy with molecular weight more than 1000
c) Compound lipophilicity	c) Affects the nose to bllood and nose to brain absorption
d) P <sup>H</sup> and pka	d) Unionized ph favours for absorption

**i) Low bioavailability**

Lipophilic drugs are generally well absorbed from the nasal cavity compared to polar drugs. The pharmacokinetic profiles of lipophilic drugs are often identical to those obtained after intravenous injection and bioavailability approaching 100%. A good examples of this is the nasal administration of Fentanyl where the *t*<sub>max</sub> for both intravenous and nasal administration have been shown to be very rapid (7 min or less) and the bioavailability for nasal anterior part of the nasal cavity can decrease clearadministration was near 80% .

**Strategies to improve nasal bioavailability:**

1. Nasal Enzyme Inhibitors  
e.g. bestatin, amastatin, borolucine, fusidic acids and bile salts
2. Nasal permeation enhancers  
e.g. Cyclodextrins, surfactants, saponins ,fusidic acids and phospholipids
3. Prodrug approach  
e.g.cyclic prodrugs, esters and derivatisation of C and N termini
4. Nasal mucoadhesives  
e.g. carbopol, polycarbophil, cellulose derivatives, lecithin and chitosan
5. Particulate drug delivery  
e.g. microspheres, nanoparticles and liposomes

**ii) Low membrane transport**

Particles entrapped in the mucus layer are transported with it and thereby effectively cleared from the nasal cavity. The combined action of the mucus layer and cilia is called mucociliary clearance. This is an important, non-specific physiological defence mechanism of the respiratory tract to protect against noxious inhaled materials. Mucus traps the particles of dust, bacteria and drug substances and is transported towards the nasopharynx at a speed of 5 - 8 mm/min, where it is swallowed. The normal mucociliary transit time in humans has been reported to be 12 to 15 min.

Low membrane transport is the general rapid clearance of the administered formulation from the nasal cavity due to the mucociliary clearance mechanism. This is especially the case for drugs that are not easily absorbed across the nasal membrane. It has been shown that for both liquid and powder formulations, that are not mucoadhesive, the half life of clearance is in the order of 15–20 min.

**Table 5: Pathological condition and their impact on mucociliary clearance:**

Pathological condition	Mucociliary clearance
<ul style="list-style-type: none"> <li>Primary ciliary dyskinesia</li> </ul>	Impaired: absence or dyskinetic beating cilia
<ul style="list-style-type: none"> <li>Asthma</li> </ul>	Increased: inflammatory process and irritation Decreased: epithelial damage
<ul style="list-style-type: none"> <li>Cystic fibrosis</li> </ul>	Impaired: dehydration of mucus
<ul style="list-style-type: none"> <li>Viral and bacterial infections</li> </ul>	Compromised: loss of cilia and change of mucus properties
<ul style="list-style-type: none"> <li>Diabetes mellitus</li> </ul>	Impaired: dehydration and microvascular damage

**iii) Enzymatic Degradation**

The role of the enzymatic barrier is to protect the lower respiratory airways from toxic agents; the nasal mucosa contains many enzymes such as cytochrome P450-dependent monooxygenase, carboxyl esterase and amino peptidase. Although nasal delivery avoids hepatic first-pass metabolism to some extent, the nasal mucosa provides a pseudo-first-pass effect. In addition, there are various barriers in the nasal membrane for protection from the microorganisms, allergens and irritating substances from the environment that must be overcome by drugs before they can be absorbed into the systemic circulation. Another contributing (but normally considered less important) factor to the low transport of especially peptides and proteins across the nasal membrane is the possibility of an enzymatic de-gradation of the molecule either within the lumen of the nasal cavity or during passage across the epithelial barrier. These sites both



contain exopeptidases such as mono- and diaminopeptidases that can cleave pep-tides at their N and C termini and endopeptidases such as serine and cysteine, which can attack internal pep-tide bonds .

#### iv) **Protective barriers**

The first step in the absorption of drugs from the nasal cavity passed through the mucus. Uncharged substances with small molecular weight can easily pass through this layer. However, larger or charged particles may find it more difficult to cross. Mucin, the principal protein in the mucus, has the potential to bind to solutes, hindering diffusion. Additionally, structural changes in the mucus layer are possible as a result of environmental changes such as pH, temperature etc. The nasal membrane is a physical barrier and the mucociliary clearance is a temporal barrier to drug absorption across the nasal epithelium<sup>2</sup>

#### **Physicochemical properties of drugs**

The influence of physicochemical characteristics of drug molecules on the rate and extent of gastrointestinal absorption is well understood. Therefore, *in silico* models have been developed to prioritize numerous drug candidates at the early phases of drug discovery. In same way, but with some differences, the physicochemical properties of drugs (molecular weight, lipophilicity, pKa, stability and solubility) can influence nasal absorption<sup>3</sup>

#### **Molecular weight, lipophilicity and pKa**

Lipophilic drugs such as propranolol, progesterone and fentanyl are, in general, well absorbed from the nasal cavity, presenting after intravenous administration (Figure 4) and a nasal bioavailability near to 100%. Indeed, they are quickly and efficiently absorbed across the nasal membrane through transcellular mechanisms. However, it is important to state that this is true for lipophilic compounds presenting a molecular weight lower than 1 kDa. The extension of nasal absorption of lipophilic drugs bigger than 1 kDa is significantly reduced .<sup>7</sup> On the other hand, the rate and degree of nasal absorption of polar drugs is low and highly dependent of the molecular weight. Several studies<sup>6, 7, 9</sup> demonstrated that the permeation of polar drugs with a molecular weight of less than 300 Da is not considerably influenced by their physicochemical properties. By contrast, the rate of permeation is highly sensitive to molecular size if it is higher than 300 Da; an inverse relationship exists between rate of permeation and molecular weight<sup>6, 7</sup>. For some small polar molecules only a 10% bioavailability is suggested. The value goes down to 1% for large molecules such as proteins<sup>53</sup>. The nasal membrane is predominantly lipophilic, hence, drug absorption is expected to diminish with a decrease in lipophilicity<sup>54,55</sup>. Thus, it is evident that polar drugs are not easily transported across nasal membrane thereby enhancing MCC. However, if lipophilicity is too high, the drug does not dissolve easily in the aqueous environment of nasal cavity, hence, with accelerated MCC the contact time with nasal membrane diminishes resulting in a reduced permeation through the wall<sup>56</sup>. In general, the passage across biomembranes is affected not only by lipophilicity/hydrophilicity, but also by the amount of drug existing as uncharged species. This depends on the drug pKa and the pH of the absorption site (5.0-6.5 in human nasal mucosa)<sup>57,58,59</sup>. According to pH partition theory, the non-ionized fraction of a drug is more permeable than that ionized. For the nasal mucosa, a range of studies evaluating the effect of lipophilicity and pH on the absorption of small drugs were performed<sup>60,61,62,63,64</sup>. All of them demonstrated that nasal absorption of weak electrolytes

depends on their ionization degree and the largest absorption occurs for the nonionized species. In this state, they present a higher apparent partition coefficient and, thus, they are more lipophilic. However, drugs such as acetylsalicylic acid<sup>60</sup> and benzoic acid<sup>61</sup> showed some permeability across the membrane even in environments that they are expected to exist as the ionized species. Based on these observations, it was concluded that, for polar drugs, partition coefficient is the major factor influencing the permeability through nasal pharmacokinetic profiles similar to those obtained.

### **Stability**

During the development of new drug formulations biological, chemical and physical drug stability in all process. As discussed before, the environment of nasal cavity has the ability to metabolize drugs by defensive enzymatic mechanisms, which may reduce the biological stability of nasally administered drugs<sup>67,68</sup>.

To overcome this difficulty a variety of strategies may be followed, mainly through the use of prodrugs<sup>69,70,71</sup> and enzymatic inhibitors<sup>76-79</sup> as it will be discussed later. On the other hand, many drugs may be physicochemically instable due to hydrolysis, oxidation, isomerisation, photochemical decomposition or polymerization reactions<sup>73</sup>. The same holds true during the intranasal drug delivery<sup>74</sup>.

### **Solubility**

Drug dissolution is a pre-requisite for any drug absorption, since only the molecularly dispersed form of a drug at the absorption site may cross the biomembranes. Hence, before nasal absorption the drug must be dissolved in the watery fluids of the nasal cavity. Thus, of the utmost importance is the appropriated aqueous drug solubility to allow enough contact with the nasal mucosa and posterior absorption<sup>22</sup>. However,

the absorption profile is influenced not only by drug solubility but also by the nature of pharmaceutical preparations, which have to guarantee the delivery of drug at therapeutically relevant doses. Due to the small size of nasal cavity, the allowable volume of drug solution is low for intranasal drug administration<sup>37</sup>. studies must be a matter of the major importance. Thereby, drugs poorly soluble in water and/or requiring high doses may constitute a problem. This can be overcome enhancing the drug aqueous solubility<sup>73,71,76,77</sup>.

### **Effect of drug formulation**

#### **Viscosity**

As formulation viscosity increases, the contact time between drug and nasal mucosa enhances and, thereby, the potential of drug absorption increases. At the same time, high viscosity of formulations interferes with normal ciliary beating and/or MCC and, thus, increases the permeability of drugs. This has been observed during nasal delivery of insulin<sup>58</sup>, acyclovir<sup>74</sup> and metoprolol<sup>61</sup>. However, sometimes, enhancing formulation viscosity does not enhance the drug absorption. For example, Zaki et al.<sup>22</sup> performed a study to evaluate the influence of formulation viscosity on the retention time of metoclopramide hydrochloride in nasal cavity and on its absorption. Interestingly, they observed that although the residence time enhanced as viscosity increased the drug absorption diminished. This observation has been attributed to a decrease in the drug diffusion from the formulation. On the other hand, it has also been reported that the viscosity of the solution may provide a larger therapeutic period of nasal formulations<sup>22</sup>.

**pH**

The extent of nasal absorption depends on the  $pK_a$  of drug and pH at the absorption site, contributing for that also the pH of formulation. At this point, it should be stated that the pH of formulation must be selected attending to drug stability and if possible should be assured the greatest quantity of non-ionized drug species.

However, the pH of formulation can induce nasal mucosa irritation and, hence, it should be similar to that found on human nasal mucosa (5.0-6.5) (26, 35, 120). Besides, the pH often prevents the bacteria growth<sup>33</sup>. In order to evaluate the effect of pH solution on the integrity of nasal mucosa, Pujara et al. (128) dissolved drugs in phosphate buffer at different pH values in the range of 2-12. The study was performed in rats whose nasal pH is 7.39<sup>20</sup> and the results demonstrated that when pH ranged from 3-10 minimal quantities of proteins and enzymes were released from cells, demonstrating no cellular damages. On the contrary, if pH values were below 3 or above 10 damages were observed intracellularly and at membrane level.

**Pharmaceutical form**

Nasal drops are the simplest and the most convenient nasal pharmaceutical form, but the exact amount of drug delivered is not easily quantified and often results in overdose<sup>11</sup>. Moreover, rapid nasal drainage can occur when using this dosage form. Solution and suspension sprays are preferred over powder sprays because the last one easily prompted the development of nasal mucosa irritation<sup>12</sup>. Recently, gel devices have been developed for a more accurate drug delivery. They reduce postnasal drip and anterior leakage, fixing the drug formulation in nasal mucosa. This enhances the drug residence time and diminishes MCC, thereby, potentially increases the nasal absorption. Over the last years, specialized systems such as lipid emulsions, microspheres, liposomes and films have also been developed to improve nasal drug delivery.

**Pharmaceutical excipients**

In nasal formulations, a wide variety of pharmaceutical excipients can be found and they are selected accordingly to their functions. Solubilizers, buffer components, antioxidants, preservatives, humectants, gelling/viscosifying agents, and flavoring or taste masking agents are some of the most usual excipients<sup>11</sup>. Although they are responsible for several nasal irritations, antioxidants, preservatives, humectants and flavoring or taste masking agents are not expected to alter nasal drug absorption<sup>11</sup>. Commonly used excipients that are frequently added to nasal preparations can be listed as below:

**Bioadhesive polymers:** It can be defined as a compound that is capable of interacting with biological material through interfacial forces and being retained on such material for prolonged periods of time. If the biological material is a mucus membrane, the bioadhesive material is termed as a mucoadhesive.<sup>4</sup>

**Examples:**

- a) Carbopol (carboxy polyethylene)
- b) Sodium carboxy methyl cellulose (SCMC)
- c) Hydroxypropyl cellulose (HPC)
- d) Hydroxypropyl methyl cellulose (HPMC)
- e) Hydroxyl ethyl cellulose (HEC)
- f) Methyl cellulose (MC)

- g) Sodium hyaluronate
- h) Guar gum
- i) Sodium alginate
- j) Polycarbophil
- k) Starch
- l) Dextran

**Gelling agent:**

Increasing solution viscosity may provide a means of prolonging the therapeutic effect of nasal preparations. A drug carrier such as hydroxypropyl cellulose was effective for improving the absorption of low molecular weight drugs but did not produce the same effect for high molecular weight peptides.

**Penetration enhancer:<sup>9</sup>**

Unlike the most small drug molecules, some drugs and peptides do not cross the nasal membrane efficiently. As a result the nasal bioavailability in simple solution formulation is very low. The low nasal absorption can be attributed to poor membrane permeability due to molecular size, lack of lipophilicity or enzymatic degradation. Enzyme inhibitors can be added to nasal formulation to prevent enzymatic degradation. The nasal mucosa is almost impermeable to molecular size greater than 1000 Dalton. To overcome these problems of poor membrane permeability most frequent used approach is the use of absorption enhancers.

They act by one or combination of the following mechanisms:

1. Alteration of properties of mucosa layer.
2. Opening tight junctions between epithelial cells.
3. Reversed micelle formation between membranes.
4. Increasing the membrane fluidity by,
  - a) Extraction or leaching of membrane components.
  - b) Creating disorders in the phospholipids domain in the membrane.

**Various types of penetration enhancers<sup>15</sup>:**

Surfactants, Bile salts, Chelators, Phospholipids, Cyclodextrins, Polyoxyethylene-9-lauryl ether (BL-9) in saline solution improves the nasal absorption of hydralazine in both in-situ and in vivo nasal absorption studies in rats. Most peptides and proteins show insufficient nasal bioavailability. Number of approaches has been described to improve their systemic bioavailability.

**Strategies for improving drug availability in nasal administration:**

- 1) To improve the nasal residence time.
- 2) To enhance the nasal absorption.
- 3) To modify drug structure to change physicochemical properties.

**Buffers:**

Nasal formulations are generally administered in small volumes ranging from 25 to 200  $\mu$ L with 100  $\mu$ L being the most common dose volume. Hence, nasal secretions may alter the pH of the administered dose. This can affect the concentration of un-ionized drug available for absorption. Therefore, an adequate formulation buffer capacity may be required to maintain

**Nasal pH**

- Nasal secretion of adult : 5.5-6.5
- Infants and children: 5-6.7

**Solubilizers:**

Aqueous solubility of drug is always a limitation for nasal drug delivery insolution. Conventional solvents or co-solvents such as glycols, small quantities of alcohol, Transcutol ( diethylene glycol monoethyl ether), medium chain glycerides and Labrasol(saturated polyglycolized C8- C10 glyceride) can be used to enhance the solubility of drugs.

**Preservatives:**

Most nasal formulations are aqueous based and need preservatives to prevent microbial growth. Parabens, benzalkonium chloride, phenyl ethyl alcohol, EDTA and benzoyl alcohol are some of the commonly used preservatives in nasal formulations. Mercury containing preservatives have a fast and irreversible effect on ciliary movement and should not be used in nasal systems.

**Antioxidants:**

A small quantity of antioxidants may be required to prevent drug oxidation. Commonly used antioxidants are sodium metabisulfite, sodium bisulfite, butylated hydroxytoluene and tocopherol.

**Humectants:**

Many allergic and chronic diseases are often connected with crusts and drying of mucous membrane. Certain preservatives/ antioxidants among other excipients are also likely to cause nasal irritation especially when used in higher quantities. Humectants avoid nasal irritation and are not likely to affect drug absorption. Common examples include glycerin, sorbitol and mannitol.

**Surfactants:**

Incorporation of surfactant into nasal dosage forms could modify the permeability of nasal membranes, which may facilitate the nasal absorption of drug.  
eg. Sodium taurocholate, sodium glycolate, polysorbate 80, sodium lauryl sulphate.

**REFERENCES:**

1. S. Upadhyay, A. Parikh, P. Joshi, U M Upadhyay and N P Chotai , Intranasal drug delivery system- A glimpse to become Maestro Journal of Applied Pharmaceutical Science 01 (03); 2011: 34-44.
2. S. Basu and A.K. Bandyopadhyay, Intranasal drug delivery :an overview Int J Pharm Sci Tech Vol-4, 2010 ISSN:0975-0525.
3. A. Pires, A. Fortuna, G. Alves and A. Falcão , Intranasal Drug Delivery: How, Why and What for?, J Pharm Pharmaceut S (www.cspsCanada.org) 12(3) 288 - 311, 2009 288.
4. U.R. Lokhande, V.D. Gorde, M.V. Gadhave., S.T. Tambe Nasal drug delivery system: a review International Standard Serial Number (ISSN): 2249-6793 International Journal of Universal Pharmacy and Life Sciences 2(2): 2012
5. D. Bhowmik, R. Kharel, J. Jaiswal, Chiranjib, Biswajit and K. P. Sampath Kumar. Innovative approaches for nasal drug delivery system and its challenges and opportunities, Annals of Biological Research, 2010, 1 (1): 21-26.
6. H. Parmar, S. Bakliwal, N. Gujarathi, B. Rane, S. Pawar. Different methods of formulation and evaluation of Mucoadhesive microsphere Volume: I: Issue-3: 2010.

7. C. McMartin. Analysis of structural requirements for the absorption of drugs and macromolecules from the nasal cavity J Pharm Sci, 1987; 76:535-540.
8. DC Corbo . Characterization of the barrier properties of mucosal membranes J Pharm Sci, 1990; 79:202-206.
9. R. Krishnamoorthy, A.K. Mitra, Prodrugs for nasal drug delivery. Advanced Drug Delivery Reviews1998; 29: 135–146, 1998.
10. YW Chien, SF Chang . Intranasal drug delivery for sys-temic medications. Crit Rev Ther Drug Carr Syst 1987;4:67-194.
11. K. R. Jadhav,M. N. Gambhire, I. M. Shaikh, V. J. Kadam and S. S. Pisal, Nasal Drug Delivery System-Factors Affecting and Applications, Current Drug Therapy, 2007, 2, 27-38 .
12. M. Rathananand, D. S. Kumar, A. Shirwaikar, R. kumar, D. Sampath kumar, Preparation of Mucoadhesive Microspheres for Nasal Delivery by Spray Drying, Indian Journal of Pharmaceutical Sciences, 2007,652.
13. P.H.Johnson, S.C.Quay, 2005. Advances in nasal drug delivery through tight junction technology. Expert Opin. Drug Deliv. 2, 281–298.
14. M.E. Aulton “Pharmaceutics –The science of dosage form design” Churchill Livingston., 494, 2002,
15. S.S.Kadam., K.R.Mahadik, A.P.Pawar, A.R.Paradkar, Transnasal delivery of peptides – a review, The East. Pharm. 1993, 47.
16. N. K. Jain (1997). Controlled and Novel Drug Delivery, Mucoadhesive drug delivery. First edition, 353.
17. S.Hirai, T.Yashiki, H.Mima, Effect of surfactants on nasal absorption of insulin in rats, Int. J. Pharm., 1981,9, 165-171.;
18. L.Illum . Nasal drug delivery: possibilities, problems and solutions. J Control Release2003; 87:187-198.
19. A.Stevens ,J, Lowe , Human histology,Mosby, Philadelphia, USA, 1997.
20. FW Merkus , JC Verhoef , NG Schipper, E Marttin . Nasal mucociliary clearanceas a factor in nasal drug delivery. Adv Drug Deliv Rev, 1998; 29:13-38.
21. JS Kimbell , EA Gross , RB Richardson ,RB Conolly , KT Morgan . Correlation of regional formaldehyde flux predictions with the distribution of formaldehyde-induced squamous metaplasia in F344 rat nasalpassages. Mutat Res, 1997; 380:143-154.
22. M.Gosau , D.Rink , O Driemel , FG Draenert . Maxillary sinus anatomy: a cadaveric study with clinical implications. Anat Rec, 2009;292:352-354.
23. YW Chien , SF Chang . Intranasal drug delivery for systemic medications. Crit Rev Ther Drug Carrier Syst, 1987; 4:67-194.
24. P Dondeti ,H Zia , TE Needham . Bioadhesive and formulation parameters affecting nasal absorption. Int J Pharm, 1996; 127:115-133.
25. P Verdugo . Goblet cells secretion andmucogenesis Annu Rev Physiol, 1990; 52:157-176.
26. Dae-Duk Ki, Drug Absorption Studies: In situ, In vitro and In silico models, chapter 9Springer, USA, 2007.
27. BJ Lipworth , CM Jackson . Safety of inhaled and intranasal corticosteroids: lessons for the new millennium. Drug Saf, 2000; 23:11-33.
28. U Baumann . Mucosal vaccination against bacterial respiratory infections. Expert Rev Vaccines, 2008; 7:1257-1276.
29. S Charlton , NS Jones, SS Davis, L Illum . Distribution and clearance of bioadhesive formulations from the olfactory region in man: Effect of polymer type and nasal delivery device. Eur J Pharm Sci, 2007; 30:295-302.
30. A. Stevens , J.Lowe , Human histology,Mosby, Philadelphia, USA, 1997.
31. VD Romeo, J Meireles, AP Sileno, HK Pimplaskar, CR Behl . Effects of physicochemical properties and other factors on systemic nasal delivery. Adv Drug Deliv Rev, 1998; 29:89-116.
32. T Furubayashi ,A Kamaguchi, K Kawaharada , Y Masaoka , M Kataoka , S Yamashita, Y Higashi, T Sakane . Evaluation of the Contribution of the Nasal Cavity andGastrointestinal Tract to Drug Absorption Following Nasal Application to Rats. Bio Pharm Bull, 2007; 30:608-611.
33. AK Leonard , AP Sileno , GC Brandt , CA Foerder , SC Quay , HR Costantino . In vitro formulation optimization of intranasal galantamine leading to enhanced bioavailabilityand reduced emetic response invivo. Int J Pharm, 2007; 335:138-146.
34. A Heidari , H Sadrai , J Varshosaz . Nasal delivery of insulin using bioadhesive chitosangels. Drug Delivery, 2006; 13:31-38.



35. MI Ugwoke , G Kaufmann , N Verbeke , R Kinget . Intranasal bioavailability of apomorphinem from carboxymethylcellulose-based drug delivery systems. *Int J Pharm*, 2000; 202:125- 131.
36. L Illum . Nasal drug delivery: new developments and strategies. *Drug Discov Today*, 2002; 7:1184-1189.
37. DG Stoke, KR Reber, LS Waltzman, C Erns, D Hamilton , D Gawareck , F Mermelstein , E McNicol , C Wright , DB Carr . Analgesic efficacy and safety of morphine-chitosan nasal solution in patients with moderate to severe pain following orthopedic surgery *Pain Med*, 2008; 9:3-12.
38. N Kilian , DG Müller . The effect of a viscosity and an absorption enhancer on the intra nasal Carbopol-based gels for nasal delivery of progesterone. *AAPS Pharm Sci Tech*, 2008;9:1078-1082.
39. WX Ding, XR Qi , Q Fu , HS Piao . Pharmacokinetics and pharmacodynamics of sterylglucoside-modified liposomes for levonorgestrel delivery via nasal route. *Drug Deliv*, 2007; 14:101-104.
40. Z Shao, GB Park , R Krishnamoorthy , AK Mitra . The physicochemical properties, plasma enzymatic hydrolysis, and nasal absorption of acyclovir and its 2'-ester prodrugs. *Pharm Res*, 1994; 11:237-242.
41. C Yang , H Gao, AK Mitra . Chemical stability, enzymatic hydrolysis, and nasal uptake of amino acid ester prodrugs of acyclovir. *J Pharm Sci*, 2001; 90:617-624.
42. IA Alsarra, AY Hamed , FK Alanazi. Acyclovir liposomes for intranasal systemic delivery: development and pharmacokinetics evaluation. *Drug Deliv*, 2008; 15:313-321.
43. S Yu, Y Zhao, F Wu, X Zhang , W Lü , H Zhang, Q Zhang . Nasal insulin delivery in the chitosan solution: in vitro and in vivo studies. *Int J Pharm*, 2004; 281:11-23.
44. J Varshosaz , H Sadrai, A Heidari . Nasal delivery of insulin using bioadhesive chitosan gels. *Drug Deliv*, 2006; 13:31-38.
45. J Wang , Tabata Y, Morimoto K. Aminated gelatin microspheres as a nasal delivery system for peptide drugs: Evaluation of in vitro release and in vivo insulin absorption in rats. *J Control Release*, 2006; 113:31-37.
46. Karasulu E, Yavasolu A, Evrensanal Z, Uyanikgil Y, Karasulu HY. Permeation studies and histological examination of sheep nasal mucosa following administration of different nasal formulations with or without absorption enhancers. *Drug Deliv*, 2008; 15:219-225.
47. Onischuk AA, Tolstikova TG, Sorokina IV. Anti-inflammatory effect from indomethacin nanoparticles inhaled by male mice. *J Aerosol Med Pulm Drug Deliv*, 2008; 21:231- 243.
48. Leykin Y, Casati A, Rapotec A. A prospective, randomized, double-blind comparison between parecoxib and ketorolac for early postoperative analgesia following nasal surgery. *Minerva Anesthesiol*, 2008; 74:475-479.
49. Moodie JE, Brown CR, Bisley EJ. The safety and analgesic efficacy of intranasal ketorolac in patients with postoperative pain. *Anesth Analg*, 2008; 107:2025-2031.
50. Romeo VD, Meireles J, Sileno AP, Pimplaskar HK, Behl CR. Effects of physicochemical properties and other factors on systemic nasal delivery. *Adv Drug Delivery Rev*, 1998; 29:89-116.
51. Costantino HR, Illum L, Brandt G, Johnson PH, Quay SC. Intranasal delivery: Physicochemical and therapeutic aspects. *Int J Pharm*, 2007; 337:1-24.
52. Leonard AK, Sileno AP, Brandt GC, Foerder CA, Quay SC, Costantino HR. In vitro formulation optimization of intranasal galantamine leading to enhanced bioavailability and reduced emetic response in vivo. *Int J Pharm*, 2007; 335:138-146.
53. Katdare A., Chaubal M.V., *Excipient Development for Pharmaceutical Biotechnology and Drug Delivery Systems*. Taylor & Francis Group, LLC, USA, 2006.
54. Corbo DC. Drug absorption through mucosal membranes: effect of mucosal route and penetrant hydrophilicity. *Pharm Res*, 1989; 6:848-852.
55. Donovan M, Flynn G, Amidon G. Absorption of polyethylene glycols 600 through 2000: the molecular weight dependence of gastrointestinal and nasal absorption. *Pharm Res*, 1990; 7:863-868.
56. Lipworth BJ, Jackson CM. Safety of inhaled and intranasal corticosteroids: lessons for the new millennium. *Drug Saf*, 2000; 23:11-33.
57. Wynsberghe D.V., Noback R.C., Carola R., *Human anatomy and physiology*, McGraw-Hill Companies, UK, 1994.
58. Dae-Duk Ki, *Drug Absorption Studies: In situ, In vitro and In silico models*, chapter 9, Springer, USA, 2007.
59. Washington N, Steele RJ, Jackson SJ, Bush D, Mason J, Gill DA, Pitt K, Rawlins DA. Determination of baseline human nasal pH and the effect of intranasally administered buffers. *Int J Pharm*, 2000; 198:139-146.

60. Hirai S, Yashiki T, Matsuzawa T, Mima H. Absorption of drugs from the nasal mucosa of rats. *Int J Pharm*, 1981; 7:317-325.
61. Hussain A.A., Bawarshi-Nassar R., Huang C.H., *Transnasal Systemic Medications*. Elsevier, Amsterdam, 1985.
62. Zaki NM, Awad GA, Mortada ND, Abd ElHady SS. Rapid-onset intranasal delivery of metoclopramide hydrochloride. Part I. Influence of formulation variables on drug absorption in anesthetized rats. *Int J Pharm*, 2006; 327:89-96.
63. Yang C, Gao H, Mitra AK. Chemical stability, enzymatic hydrolysis, and nasal uptake of amino acid ester prodrugs of acyclovir. *J Pharm Sci*, 2001; 90:617-624.
64. Corbo DC. Characterization of the barrier properties of mucosal membranes. *J Pharm Sci*, 1990; 79:202-206.
65. Corbo DC. Drug absorption through mucosal membranes: effect of mucosal route and penetrant hydrophilicity. *Pharm Res*, 1989; 6:848-852.
66. Donovan M, Flynn G, Amidon G. Absorption of polyethylene glycols 600 through 2000: the molecular weight dependence of gastrointestinal and nasal absorption. *Pharm Res*, 1990; 7:863-868.
67. Bogdanffy MS. Biotransformation enzymes in the rodent nasal mucosa: The value of a histochemical approach. *Environ Health Perspect*, 1990; 85:177-186.
68. Mitra AK, Krishnamoorthy R. Prodrugs for nasal drug delivery. *Adv Drug Deliv Rev*, 1998; 29:135-146.
69. Arora P, Sharma S, Garg S. Permeability issues in nasal drug delivery. *Drug Discovery Today*, 2002; 7:967-975.
70. Costantino HR, Illum L, Brandt G, Johnson PH, Quay SC. Intranasal delivery: Physicochemical and therapeutic aspects. *Int J Pharm*, 2007; 337:1-24.
71. Huang CH, Kimura R, Nassar RB, Hussain A. Mechanism of nasal absorption of drugs. I: Physicochemical parameters influencing the rate of in situ nasal absorption of drugs in rats. *J Pharm Sci*, 1985; 74:608-611.
72. Costantino HR, Illum L, Brandt G, Johnson PH, Quay SC. Intranasal delivery: Physicochemical and therapeutic aspects. *Int J Pharm*, 2007; 337:1-24.
73. Romeo VD, Meireles J, Sileno AP, Pimplaskar HK, Behl CR. Effects of physicochemical properties and other factors on systemic nasal delivery. *Adv Drug Delivery Rev*, 1998; 29:89-116.
74. Arora P, Sharma S, Garg S. Permeability issues in nasal drug delivery. *Drug Discovery Today*, 2002; 7:967-975.
75. Kao HD, Traboulsi A, Itoh S, Dittert L, Hussain A. Enhancement of the systemic and CNS specific delivery of L-dopa by the nasal administration of its water soluble prodrugs. *Pharm Res*, 2000; 17:978-984.
76. Machida M. Effects of surfactants and protease inhibitors on nasal absorption of recombinant human granulocyte colony stimulating factor (rhG-CSF) in rats. *Biol Pharm Bull*, 1994; 17:1375-1378.
77. Morimoto K, Miyazaki M, Kakemi M. Effects of proteolytic enzyme inhibitors on nasal absorption of salmon calcitonin in rats. *Int J Pharm*, 1995; 113:1-8.
78. Bernkop-Schnurch A. Use of inhibitory agents to overcome the enzymatic barrier to perorally administered therapeutic peptides and proteins. *J Control Release*, 1998; 52:1-16.